

# **TECHNICAL DATA SHEET**

# Article No. 9881

Letheen Modified Broth, ready-to-use culture medium, bottles

# **SYNONYMS**

Letheen Broth with Tween, Modified

# **SPECIFICATION**

Prepared medium. Liquid medium for the primary recovery of stressed microorganisms in the microbial examination of cosmetics according to FDA and ISO.

Colour: Yellowish-brown pH:  $7.2 \pm 0.2$  at 25 °C

## **COMPOSITION IN G/L**

| Casein peptone   | 15.0 |
|------------------|------|
| Meat peptone     | 10.0 |
| Meat extract     | 5.0  |
| Yeast extract    | 2.0  |
| Lecithin         | 0.7  |
| Sodium chloride  | 5.0  |
| Polysorbate 80   | 5.0  |
| Sodium bisulfite | 0.1  |
|                  |      |

## **PACKAGE DETAILS**

9881-10x90ML

 $\begin{array}{lll} \mbox{Volume} & 90 \pm 3 \mbox{ ml} \\ \mbox{Bottle size} & 125 \mbox{ ml} \\ \mbox{Packaging unit} & 10 \mbox{ bottles} \end{array}$ 

1 box with 10 x 90 ml in 125-ml-bottles. Injectable cap: Plastic screw inner cap + protective outer blue cap. For the use of syringe needles with a diameter  $\leq$  0.8 mm.





## 9881-10x450ML

Volume $450 \pm 5 \text{ ml}$ Bottle size500 mlPackaging unit10 bottles

1 box with 10 x 450 ml in 500-ml-bottles. Injectable cap: Plastic screw inner cap + protective outer blue cap. For the use of syringe needles with a diameter  $\leq$  0.8 mm.

#### DESCRIPTION

The use of lecithin and polysorbates to neutralise the antimicrobial effect of quaternary ammonium compounds (QACs) originates from the recommendation of Weber and Black in the 1940s.

The AOAC accepted the methodology for antimicrobial testing in 1965 and extended its application to all cationic surfactants (detergents). The TAT (tryptone azolectin polysorbate) medium in the Newburg Cosmetic Analysis Manual (2nd edition, 1977) has a similar composition and uses the AOAC formulation. The FDA (Bacteriological Analytical Manual, 5th edition, 1978) has included it as the primary presumptive and enrichment medium used for all microbial testing of cosmetics.

The present formulation appears in the 8th edition (1998) of the BAM and the notable modification are the inclusion of sodium chloride providing suitable osmotic pressure and an increased amount of peptones and tissue extracts to promote good growth. These transform the medium into a very rich all-purpose medium suitable for neutralizing almost all preservatives present in samples under examination.

The ISO Technical Committee on Cosmetics (ISO/TC 217) (2006) has also adopted the present formulation as an alternative enrichment medium prior to microbiological examination.

## **TECHNIQUE**

Collect, dilute and prepare samples and volumes as required according to specifications, directives, official standard regulations and/or expected results. Dispense liquid medium in appropriate containers if the original container is of large volume.

Inoculate aseptically the bottles/tubes with the prepared sample or its dilution.

Incubate the tubes tightly closed aerobically at 30-35 °C for 24-48-72 h

(Incubation times, temperature and sample volumes may vary depending on sample and specifications) Read turbidity increase as growth indicator.

This medium may be used to inoculate any confirmatory, secondary medium by streaking methodology or by spiral method; after proper incubation, enumerate all colonies that have appeared on the surface of the secondary agar.

Evaluation of results according to the laboratorie's specifications.

Calculate total microbial count per ml of sample by multiplying the average number of colonies per plate by the inverse dilution factor (in case of streaked diluted sample). Report results as colony forming Unit (CFU's) per ml or along with enrichment and secondary media used, incubation time and temperature.





## **MICROBIOLOGICAL CONTROL**

Prepare tubes - inoculate with 100±20 CFU for growth promotion or 10<sup>4</sup>-10<sup>6</sup> CFU (selectivity). Aerobiosis. Incubation at 30-35 °C. Reading after 24-48h until 72 h

| Microorganism                                  | Growth |
|--|--------|
| Escherichia coli ATCC® 25922, WDCM 00013       | Good   |
| Ps. aeruginosa ATCC® 9027, WDCM 00026          | Good   |
| Staphylococcus aureus ATCC® 6538, WDCM 00032   | Good   |
| Bacillus subtilis ATCC® 6633, WDCM 00003       | Good   |
| Salmonella typhimurium ATCC® 14028, WDCM 00031 | Good   |

#### Sterility control:

Incubation 48 hours at 30-35 °C and 48 hours at 20-25 °C: NO GROWTH. Check 7 days after incubation under same conditions.

# **REFERENCES**

- ASTM Standard E 640-78 (1991) Test Method for the preservatives in water-containing cosmetics. Philadelphia.
  PA. USA.
- · ATLAS, R.M., L.C. PARKS (1993) Handbook of Microbiological Media. CRC Press, Inc. London.
- · FDA (Food and Drug Adminstrations) (1998) Bacteriological Analytical Manual 8th ed. Revision A. AOAC International. Gaithersburg, MD, USA.
- · HORWITZ, W. (2000) Official Methods of Analysis. AOAC International. Gaithersburg. MD. USA.
- . ISO 11133:2014/ Adm 1:2018. Microbiology of food, animal feed and water. Preparation, production, storage and performance testing of culture media.
- · ISO 16212 Standard (2017) Cosmetics Microbiology Enumeration of yeast and mould.
- · ISO 21149 Standard (2017) Cosmetics Microbiology Enumeration and detection of aerobic mesophilic bacteria.
- · ISO 21150 Standard (2015) Cosmetics Microbiology Detection of Escherichia coli.
- · ISO 22717 Standard (2015) Cosmetics Microbiology Detection of Pseudomonas aeuruginosa.
- · ISO 22718 Standard (2015) Cosmetics Microbiology Detection of Staphylococcus aureus.
- LUCAS, I.P. (1977) Microbiological Examination of Cosmetics. Newburger's Manual of Cosmetic Analysis AOAC. Washington.
- US PHARMACOPOEIA (2002) <61> Microbial Limit Tests. 25th ed. US Pharmacopeial Convention. Rockville. MD. USA.
- WEBER, G.R. & L.A. BLACK (1948) Relative efficiency of quaternary inhibitors. Soap and Sanit. Chem. 24:134-139.





# **STORAGE**

8 - 25 °C

# SHELF LIFE

16 months unopened from date of manufacture

created: 19.08.2022

