Protocols & Specifications

IBI SCIENTIFIC

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An ISO 9001 Certified Company

QUICK BLUE PROTEIN STAIN

Product Number	*Size*
IB01034	**1L**

PHYSICAL SPECIFICATIONS

- Detect proteins down to 5ng
- Does NOT contain organic solvents or phosphoric acid
- Can be used for Mass Spectrometry
- Rinse with DI Water

STORAGE

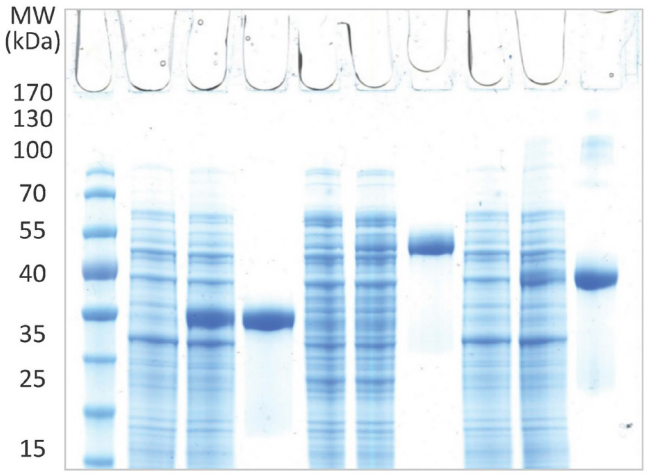
Store at room temperature for a minimum of 1 year.

CAS NO.

CAS# N/A

WARNING

Classification: No known hazards



PROTOCOL

IBI Quick Blue Protein Stain is a comprehensive rapid 1-step coomassie type stain. The proprietary formulation, using colloidal coomassie, is ideal for rapid protein staining in polyacrylamide gels. This is a non-hazardous product that does NOT utilize organic solvents or phosphoric acid. A completely environmentally friendly product!

Simple 1-Step Protocol:

1.) Pour 25ml of Quick Blue Protein Stain into a container. Use more stain if you are staining a gel larger than 10cm x 10cm or using a large container.

2.) Remove the gel from the cassette and place the gel into the stain.

3.) Using gentle/medium mixing setting on your Belly Dancer or Belly Button Shaker (or other type shaker), mix the gel for 15 minutes or until all weak protein bands are fully developed. Stain intensity is higher after about 1 - 2 hours, and is at maximum if left to incubate overnight.

4.) Transfer the gel to DI Water to remove any background staining and for gel storage. A minimum of 1 hour staining is recommended before storing gel in DI Water.

Microwave Procedure for Gels:

1.) Using a microwave to heat the Quick Blue Protein Stain can speed up the development of the protein bands.

2.) To turbo-charge the stain, we recommend microwaving the gel, immersed in the Quick Blue Protein Stain, in a suitable microwave safe container, for a maximum of 10 seconds at full power.

3.) Remove the container from the microwave and keep the gel in the stain for at least 30 minutes to 1 full hour before storing the gel in DI Water.

Mass Spectrometry Application:

1.) Stain gel as normal.

2.) Excise the protein band of interest and place inside a clean microcentrifuge tube.

3.) Add 1ml of 30% ethanol or 30% acetone.

4.) Incubate for 20 minutes (60°C - 70°C increases the rate of destaining)

5.) Decant the supernatant and repeat steps 3 and 4 at least 3 times or until the gel fragement is clear.

6.) Run the sample in the mass spectrometer.

FOR RESEARCH AND DEVELOPMENT PURPOSES ONLY

Building Quality Scientific.