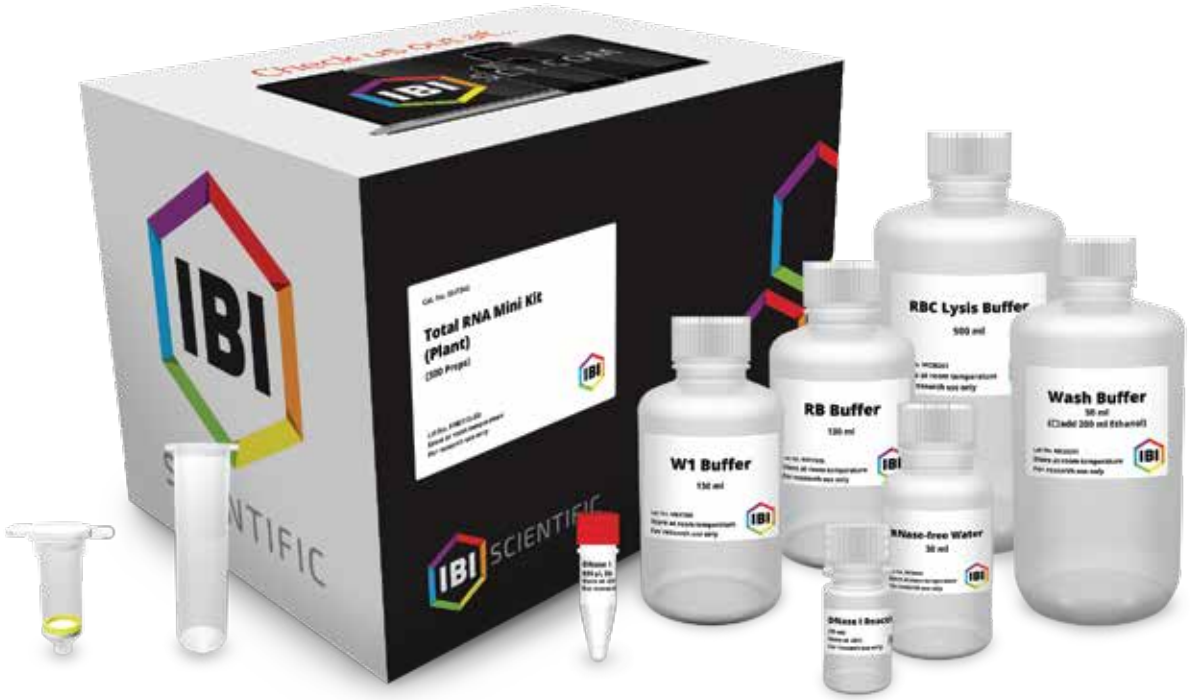
For Research Use Only



IBI SCIENTIFIC

INSTRUCTION MANUAL

Total RNA Mini Kit (Plant)

IB47340
IB47341
IB47342
IB47343

Sample: up to 100 mg of fresh plant tissue or up to 25 mg of dry plant tissue

Yield: 15-20 µg of RNA from 50 mg Arabidopsis thaliana leaf

Format: spin column

Operation Time: within 15 minutes

Elution Volume: 25-100 µl

Storage: dry at room temperature (15-25°C)

Introduction

The Total RNA Mini Kit (Plant) provides an efficient method for purifying total RNA from plant tissue and cells. Samples are ground in liquid nitrogen then filtered to remove cell debris. In the presence of a binding buffer and chaotropic salt, total RNA in the lysate binds to the glass fiber matrix of the spin column. Optional in-column DNase treatment can be followed. Once any contaminants have been removed using Wash Buffer (containing ethanol), the purified total RNA is eluted by RNase-free Water. The procedure does not require phenol extraction or alcohol precipitation and can be completed within 15 minutes. The purified total RNA is ready for use in RT, RT-PCR, Real-time PCR and Northern Blotting.

Quality Control

The quality of the Total RNA Mini Kit (Plant) is tested on a lot-to-lot basis according to IBI's ISO-certified quality management system. Total RNA is isolated from a 25 mg young leaf sample, quantified with a spectrophotometer and analyzed by electrophoresis.

Kit Contents

Component	IB47340	IB47341	IB47342	IB47343
RB Buffer	3 ml	30 ml	60 ml	160 ml
PRB Buffer	3 ml	30 ml	60 ml	160 ml
DNase I[1] (2U/µl)	20 µl	275 µl	550 µl	550 µl x 3
DNase I Reaction Buffer	200 µl	2.5 ml	5 ml	15 ml
W1 Buffer	2 ml	30 ml	50 ml	130 ml
Wash Buffer[2] (Add Ethanol)	1.5 ml (6 ml)	25 ml (100 ml)	25 ml + 12.5 ml (100 ml)(50 ml)	50 ml x 2 (200 ml x 2)
RNase-free Water	1 ml	6 ml	15 ml	30 ml
Filter Columns	4	50	100	300
RB Columns	4	50	100	300
2 ml Collection Tubes	8	100	200	600

[1] DNase I is shipped at room temperature and should be stored at -20°C for extended periods after receiving the kit.

[2] Add absolute ethanol (see the bottle label for volume) to Wash Buffer then mix by shaking for a few seconds. Check the box on the bottle. Be sure and close the bottle tightly after each use to avoid ethanol evaporation.

Order Information

Product	Package Size	Catalogue #
Total RNA Mini Kit (Blood/Cell)	50/100/300 preps	IB47321/322/323
Total RNA Mini Kit (Tissue)	50/100/300 preps	IB47301/302/303
Total RNA Mini Kit (Plant)	50/100/300 preps	IB47341/342/343
rBAC Mini RNA Bacteria Kit	100/300 preps	IB47421/422
rYeast Total RNA Mini Kit	100/300 preps	IB47411/412

Product	Package Size	Catalogue #
miRNA Isolation Kit	100 preps	IB47371
IBI Isolate	100/200 rxns	IB47601/302
IBI Tri-Isolate	100/200 rxns	IB47631/632
RNA Pure Kit	50/100 rxns	IB47641/642

Caution

RB Buffer and PRB Buffer contain chaotropic salt. During operation, always wear a lab coat, disposable gloves, protective goggles and (anti-fog) procedure mask.

Steps to prevent RNase contamination

Disposable and nondisposable plasticware and automatic pipettes should be sterile and used only for RNA procedures.

IMPORTANT BEFORE USE!

Various plant species contain different metabolites such as polysaccharides, polyphenols, and proteins. The standard protocol uses RB Buffer for lysis of most common plant species. The RB Buffer system ensures purified RNA with high yields and high quality. Alternatively, PRB Buffer is provided with the kit to ensure efficient cell lysis of plant species with high polysaccharide content.

- Add absolute ethanol (see the bottle label for volume) to Wash Buffer prior to initial use
- Additional Requirements: phosphate-buffered saline (PBS), absolute ethanol, ddH2O (RNase/DNase-free), microcentrifuge tubes, pipette tips, ß-mercaptoethanol, 0.10-0.25% Trypsin

Total RNA Mini Kit (Plant) Protocol

<table>
<tr><td>Plant Tissue Dissociation</td><td>• Cut off 50 mg (up to 100 mg) of fresh or frozen plant tissue.
• Freeze the sample with liquid nitrogen.
• Grind the sample to a fine powder then transfer it to a 1.5 ml microcentrifuge tube.
NOTE: Some plant samples can be ground sufficiently in the absence of liquid nitrogen.</td></tr>
<tr><td>Step 1 Lysis</td><td>• Add 500 µl of RB Buffer or PRB Buffer and 5 µl of ß-mercaptoethanol (or 10 µl of freshly prepared 2M Dithiothreitol in RNase Free Water) and mix by vortex.
• Incubate at 60°C for 5 minutes. Place a Filter Column in a 2 ml Collection Tube.
• Transfer the sample mixture to the Filter Column.
• Centrifuge for 1 minute at 1,000 x g then discard the Filter Column.
• Carefully transfer the clarified filtrate to a new 1.5 ml microcentrifuge tube.</td></tr>
<tr><td>Step 2 RNA Binding</td><td>• Add a 1⁄2 volume of absolute ethanol to the clarified filtrate then shake vigorously.
E.g. Add 250 µl of absolute ethanol to 500 µl of filtrate.
• Place a RB Column in a 2 ml Collection Tube then transfer the mixture to the RB Column.
• Centrifuge at 14-16,000 x g for 1 minute.
NOTE: If the mixture could not flow past the RB Column membrane following centrifugation, increase the centrifuge time until it passes completely.
• Discard the flow-through then place the RB Column back in the 2 ml Collection Tube.
Optional Step 1: In Column DNase I Digestion
The amount of DNA contamination is significantly reduced following In Column DNase I Digestion. However, traces of residual DNA may be detected in very sensitive applications. In this situation, please perform Optional Step 2: DNA Digestion In Solution instead to efficiently remove trace amounts of DNA. Standard DNase buffers are incompatible with In Column DNase I Digestion and may affect RNA integrity and reduce yield.
1. Add 400 µl of Wash Buffer (make sure ethanol was added) to the RB Column then centrifuge at 14-16,000 x g for 30 seconds.
2. Discard the flow-through and place the RB Column back in the 2 ml Collection Tube.
3. Prepare DNase I solution in a 1.5 ml microcentrifuge tube (RNase-free) as follows:
<table>
<tr><td>DNase I</td><td>5 µl (2 U/µl)</td></tr>
<tr><td>DNase I Reaction Buffer</td><td>45 µl</td></tr>
<tr><td>Total Volume</td><td>50 µl</td></tr>
</table>
4. Gently pipette DNase I solution to mix (DO NOT vortex) then add DNase I solution (50 µl) into the CENTER of the RB column matrix.
5. Incubate the column for 15 minutes at room temperature (20-30°C) then proceed with Step 3 RNA Wash.</td></tr>
<tr><td>Step 3 RNA Wash</td><td>• Add 400 µl of W1 Buffer into the center of the RB Column.
• Centrifuge at 14-16,000 x g for 30 seconds.
• Discard the flow-through then place the RB Column back in the 2 ml Collection Tube.
• Add 600 µl of Wash Buffer (make sure ethanol was added) to the center of the RB Column.
• Centrifuge at 14-16,000 x g for 30 seconds.
• Discard the flow-through then place the RB Column back in the 2 ml Collection Tube.
• Add 600 µl of Wash Buffer (make sure ethanol was added) to the center of the RB Column.
• Centrifuge at 14-16,000 x g for 1 minute.
• Discard the flow-through then place the RB Column back in the 2 ml Collection Tube.
• Centrifuge at 14-16,000 x g for 3 minutes to dry the column matrix.</td></tr>
<tr><td>Step 4 RNA Elution</td><td>• Place the dried RB Column in a clean 1.5 ml microcentrifuge tube.
• Add 50 µl of RNase-free Water to the CENTER of the column matrix.
• Let stand for at least 2 minutes to ensure the RNase-free Water is completely absorbed.</td></tr>
</table>

<table>
<tr><td>Step 4
RNA Elution</td><td>• Centrifuge at 14-16,000 x g for 1 minute to elute the purified RNA.
NOTE: If higher RNA concentration is required, repeat Step 4 using the final eluate.
Optional Step 2: DNA Digestion In Solution
1. Prepare DNase I reaction in a 1.5 ml microcentrifuge tube (RNase-free) as follows:
<table><tr><td>RNA in RNase-free Water</td><td>1 - 40 µl</td></tr><tr><td>DNase I</td><td>0.5 µl/µg RNA</td></tr><tr><td>DNase I Reaction Buffer</td><td>5 µl</td></tr><tr><td>RNase-free Water</td><td>Add to final volume = 50 µl</td></tr><tr><td>Total Volume</td><td>50 µl</td></tr></table>2. Gently pipette the DNase I reaction solution to mix (DO NOT vortex) then incubate the microcentrifuge tube at 37°C for 15-30 minutes.
3. Stop the reaction by adding 1 µl of 20 mM EGTA (pH=8.0) then incubate the microcentrifuge tube at 65°C for 10 minutes.
NOTE: DNase I Reaction Buffer may cause aberrant migration or smearing of RNA on gels. If analyzing RNA by gel electrophoresis, repurify the RNA sample by using the RNA Pure Kit instead of stopping the reaction with EGTA.</td></tr>
</table>

Troubleshooting

Problem	Possible Reasons/Solution
Clogged Column	• Insufficient disruption and/or homogenization • Too much starting material • Centrifugation temperature was too low (should be 20°C to 25°C)
Low RNA Yield	• Insufficient disruption and/or homogenization • Too much starting material • RNA still bound to RB Column membrane • Ethanol carryover
RNA Degradation	• Harvested sample not immediately stabilized • Inappropriate handling of starting material • RNase contamination

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